

**Final Laboratory Testing Report
for
Operation Phase Whole Effluent Toxicity Test for
Advance Disinfection Facilities at Stonecutters Island
Sewage Treatment Works
(Contract No. DE/2012/08)**

Report for the Month of July 2014

**R&D Corp
The Hong Kong University of Science and Technology**

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1. Introduction

1.1. Background

The whole effluent toxicity tests (WETT) were carried out under the requirements of Drainage Service Department (DSD).

1.2. Testing laboratory and investigator

The following tests were carried out in the Coastal Marine Laboratory (CML), Hong Kong University of Science and Technology.

Principle investigator: Prof. Wen-Xiong WANG

Phone number: (852) 2358-7346; **Fax number:** (852) 2358-1559

Address: Division of Life Science, the Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong

1.3. Sample

A 24-hour flow-weighted composite effluent sample was collected from Stonecutters Island Sewage Treatment Works (SCISTW) on July 22th, 2014. Effluent sample was shipped immediately to the testing laboratory on the same day of collection and stored at 4 °C until use. Toxicity testings were started on the same day after sample collection.

1.4. Test species

The following test species were included in the WETT. The acute toxicity test for amphipod (*Melita longidactyla*) was not included because of unavailability of the organisms in this month. :

- Copepod (*Tigriopus japonicus*)
- Fish (*Lutjanus malabaricus*)
- Barnacle larvae (*Balanus amphitrite*)
- Diatom (*Skeletonema costatum*)
- Shrimp (*Metapenaeus ensis*)

1.5. Test protocols

The WETT testing methods and procedures follow those documented in "Consultancy Study on Fisheries and Marine Ecological Criteria for Impact Assessment-Final Report" commissioned by Agriculture, Fisheries and Conservation Department (AFCD), as indicated in tender addendum No. 1 by Drainage Services Department (DSD).

The report is certified by:

_____ (Prof. Wen-Xiong WANG) Date: _____

2. Report on Copepod Acute Toxicity Test

Test report

2.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 µm membrane to remove the large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

2.2. Test organism

Species	Copepod (<i>Tigriopus japonicus</i>).
Source:	Obtained from the laboratory cultures
Size/age:	Adult
Acclimatization:	Acclimatized in fully aerated seawater (temperature: 22±1°C, salinity: 30‰) at least 48 hours in the laboratory prior to test. Fed with microalgae <i>Isochrysis galbana</i> .

2.3. Summary of test conditions

Type of test:	Static
Duration:	48 h, 28/07/2014-30/07/2014
Dilution seawater source:	Seawater collected from a pristine site in Clear Water Bay, Sai Kung, Hong Kong
Dilution seawater pretreatment:	Filtered through 0.22 µm membrane
Testing temperature:	22±1 °C
Lighting:	Continuous
Salinity:	30‰
Testing chamber:	Pre-cleaned 50 mL glass flask
Feeding:	None
Number of organisms per replicate:	10
Replicate number:	4
Volume of test medium:	20 mL
Aeration:	Moderate shaking with shaker (15 rpm)
Reference toxicant:	CdCl ₂
Positive control:	48 h acute toxicity test
Salinity control:	Prepared with ocean salt adding into de-ionized water, salinity: 30‰

2.4. Test results

Table 1. Survival of copepods after 48 hours.

Treatment	Effluent concentration (%)	Number of living copepods after 48 hour (individuals)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	10	10	10	10	10.0	0.0
Salinity control	0	10	10	10	10	10.0	0.0
Concentration 1	6.5	10	10	10	10	10.0	0.0
Concentration 2	12.5	10	10	9	10	9.8	0.5
Concentration 3	25	10	10	10	10	10.0	0.0
Concentration 4	50	9	8	10	10	9.3	1.0
Concentration 5	100	4	6	5	4	4.8	1.0

Table 2. Survival percentage of copepods after 48 hours.

Treatment	Effluent concentration (%)	Percentage of living copepods after 48 hour (%)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	100	100	100	100	100.0	0.0
Salinity control	0	100	100	100	100	100.0	0.0
Concentration 1	6.5	100	100	100	100	100.0	0.0
Concentration 2	12.5	100	100	90	100	97.5	5.0
Concentration 3	25	100	100	100	100	100.0	0.0
Concentration 4	50	90	80	100	100	92.5	9.6
Concentration 5	100	40	60	50	40	47.5	9.6

2.5. Summary of water quality parameters monitoring during test.

Table 3. Summary of water quality parameters during copepod acute toxicity test.

Water quality parameters	Effluent concentration (%)						
	Negative control	Salinity control	6.5	12.5	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L ⁻¹)	6.8-7.0	6.7-7.2	6.7-7.0	6.7-7.2	6.7-7.0	6.7-7.2	7.0-7.2
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.8-8.0	7.8-8.1	7.8-8.0	7.8-8.1	7.8-8.1	7.9-8.1	7.9-8.1
Total ammonia (start/end, mg L ⁻¹)	0.03/0.13	0.07/0.12	0.92/0.83	1.81/2.04	3.91/4.11	7.72/7.48	14.3/13.2
Total sulfide (start/end, mg L ⁻¹)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Total residual chlorine (start/end, mg L ⁻¹)	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02
Total suspended solid (start/end, mg L ⁻¹)	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2

2.6. LC₅₀ for the copepod *Tigriopus japonicus* and test acceptability

Table 4. LC₅₀ for the copepods and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	(94.7±2.2)%	NA
Negative control survival	100%	>90%
Reference toxicant 48-h acute test	4.6 mg L ⁻¹	4.3±0.5 mg L ⁻¹
95% of confidence range of reference toxicant test	4.1-6.0 mg L ⁻¹	NA
Daily temperature variation	<0.1 °C	Average daily temperature variation: ±1 °C
Dissolved oxygen concentration	>6.7 mg L ⁻¹	>4 mg L ⁻¹

NA: Not applicable

3. Report on Fish Acute Toxicity Test

Test report

3.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

3.2. Test organism

Species	Fish (<i>Lutjanus malabaricus</i>)
Source:	Purchased from local contracted fish farm
Size/age:	5-8 cm
Acclimatization:	Acclimatized in fully aerated seawater (temperature: 22±1°C, salinity: 30‰) at least 48 hours in laboratory prior to test. Fed with fresh shrimp purchased from local market.

3.3. Summary of test conditions

Type of test:	Static
Duration:	48 h, 28/07/2014-30/07/2014
Dilution seawater source:	Seawater collected from a pristine site in Clear Water Bay, Sai Kung, Hong Kong
Dilution seawater pretreatment:	Filtered through 5 µm filtration bag
Testing temperature:	22±1 °C
Lighting:	Continuous
Salinity:	30‰
Testing chamber:	Pre-cleaned 20 L tank
Feeding:	None
Number of organisms per replicate:	10
Replicate number:	4
Volume of test medium:	20 L
Aeration:	Moderate, with air stone
Reference toxicant:	CdCl ₂
Positive control:	48 h acute toxicity test
Salinity control:	Prepared with ocean salt adding into de-ionized water, salinity: 30‰

3.4. Test results

Table 1. Survival of fish after 48 hours.

Treatment	Effluent concentration (%)	Number of living fish after 48 hour (individuals)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	10	10	10	10	10.0	0.0
Salinity control	0	10	10	9	10	9.8	0.5
Concentration 1	6.5	10	9	10	10	9.8	0.5
Concentration 2	12.5	10	10	10	10	10.0	0.0
Concentration 3	25	10	10	8	10	9.5	1.0
Concentration 4	50	9	7	9	8	8.3	1.0
Concentration 5	100	1	1	0	0	0.5	0.6

Table 2. Survival percentage of fish after 48 hours.

Treatment	Effluent concentration (%)	Percentage of living fish after 48 hour (%)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	100	100	100	100	100.0	0.0
Salinity control	0	100	100	90	100	97.5	5.0
Concentration 1	6.5	100	90	100	100	97.5	5.0
Concentration 2	12.5	100	100	100	100	100.0	0.0
Concentration 3	25	100	100	80	100	95.0	10.0
Concentration 4	50	90	100	100	100	97.5	5.0
Concentration 5	100	10	10	0	0	5.0	5.8

3.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during fish acute toxicity test.

Water quality parameters	Effluent concentration (%)						
	Negative control	Salinity control	6.5	12.5	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L ⁻¹)	7.3-7.7	7.2-7.7	7.2-7.5	7.3-7.7	7.1-7.5	7.1-7.6	7.3-7.5
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.9-8.1	7.7-8.1	7.9-8.2	7.9-8.2	8.0-8.2	8.1-8.2	7.5-7.9
Total ammonia (start/end, mg L ⁻¹)	0.07-0.09	0.06/0.11	1.48/0.87	2.31/1.02	4.79/2.35	7.11/3.91	15.1/8.62
Total sulfide (start/end, mg L ⁻¹)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Total residual chlorine (start/end, mg L ⁻¹)	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02
Total suspended solid (mg L ⁻¹)	3.6/4.9	4.5/5.1	6.5/7.5	10.1/4.5	15.8/5.7	20.3/8.6	31.2/19.2

3.6. LC₅₀ for the fish *Lutjanus malabaricus* and test acceptability

Table 4. LC₅₀ for the fish and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	(70.6±1.9)%	NA
Negative survival	100.0%	> 90%
Reference toxicant 48-h acute test	13.5 mg L ⁻¹	14.4±1.8 mg L ⁻¹
95% of confidence range of reference toxicant test	13.4-15.7 mg L ⁻¹	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: ± 1 °C
Dissolved oxygen concentration	>7.1 mg L ⁻¹	> 4 mg L ⁻¹

NA: Not applicable

4. Report on Barnacle Larvae Acute Toxicity Test

Test report

4.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 µm membrane filter to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately to dissolved oxygen (DO) saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

4.2. Test organism

Species	Barnacle larvae (<i>Balanus amphitrite</i>).
Source:	Introduced from adult barnacles collected from Sai Kung
Size/age:	Stage II
Acclimatization:	Acclimatized in fully aerated seawater held in 500 mL glass beaker (temperature: 22±1°C, salinity: 30‰) for at least 24 hours in laboratory prior to test. Fed with diatom <i>Chaetoceros gracilis</i> .

4.3. Summary of test conditions

Type of test:	Static
Duration:	48 h, 03/08/2014-05/08/2014
Dilution seawater source:	Seawater collected from a pristine site in Clear Water Bay, Sai Kung, Hong Kong
Dilution seawater pretreatment:	Filtered through 0.22 µm membrane
Testing temperature:	22±1 °C
Lighting:	Continuous
Salinity:	30‰
Testing chamber:	Pre-cleaned 50 mL glass beaker
Feeding:	None
Number of organisms per replicate:	20
Replicate number:	4
Volume of test medium:	20 mL
Aeration:	Moderate, around 100 bubbles/min
Reference toxicant:	CdCl ₂
Positive control:	48 h acute toxicity test
Salinity control:	Prepared with ocean salt adding into de-ionized water, salinity: 30‰

4.4. Test results

Table 1. Survival of barnacle larvae after 48 hours

Treatment	Effluent concentration (%)	Number of living barnacle larvae after 48 hour (individuals)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	19	20	19	20	19.5	0.6
Salinity control	0	17	20	19	19	18.8	1.3
Concentration 1	6.5	17	17	18	16	17.0	0.8
Concentration 2	12.5	13	14	12	16	13.8	1.7
Concentration 3	25	11	14	11	12	12.0	1.4
Concentration 4	50	10	10	8	10	9.5	1.0
Concentration 5	100	3	2	4	0	2.3	1.7

Table 2. Survival percentage of barnacle larvae after 48 hours.

Treatment	Effluent concentration (%)	Percentage of living barnacle larvae after 48 hour (%)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	95	100	95	100	97.5	2.9
Salinity control	0	85	100	95	95	93.8	6.3
Concentration 1	6.5	85	85	90	80	85.0	4.1
Concentration 2	12.5	65	70	60	80	68.8	8.5
Concentration 3	25	55	70	55	60	60.0	7.1
Concentration 4	50	50	50	40	50	47.5	5.0
Concentration 5	100	15	10	20	0	11.3	8.5

4.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during barnacle larvae acute toxicity test

Water quality parameters	Effluent concentration (%)						
	Negative control	Salinity control	6.5	12.5	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L ⁻¹)	6.7-7.0	6.5/6.7	6.7-6.9	6.7-6.9	6.6-6.8	6.7-6.9	6.8-7.0
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.9-8.1	7.8-8.1	7.9-8.1	7.9-8.2	7.9-8.1	7.8-8.1	7.6-7.9
Total ammonia (start/end, mg L ⁻¹)	0.05/0.07	0.09/0.05	0.87/0.65	1.66/1.52	3.72/3.53	6.83/6.06	12.0/11.1
Total sulfide (start/end, mg L ⁻¹)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Total residual chlorine (start/end, mg L ⁻¹)	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02
Total suspended solid (mg L ⁻¹)	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2

4.6. LC₅₀ for the barnacle larvae *Balanus amphitrite* and test acceptability

Table 4. LC₅₀ for the barnacle larvae and test acceptability

Parameter	Value	Control limit
Calculated LC ₅₀	(47.2±4.1)%	NA
Negative survival	97.5%	>90%
Reference toxicant 48-h acute test	1.20 mg L ⁻¹	1.04±0.11 mg L ⁻¹
95% of confidence range of reference toxicant test:	0.95-1.11 mg L ⁻¹	NA
Daily temperature variation	<0.1 °C	Average daily temperature variation: ± 1 °C
Dissolved oxygen concentration	>6.5 mg L ⁻¹	> 4 mg L ⁻¹

NA: Not applicable

5. Report on Diatom Growth Inhibition Test (Chronic toxicity test)

Test report

5.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 µm membrane filter to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately to dissolved oxygen (DO) saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

5.2. Test organism

Species	Diatom (<i>Skeletonema costatum</i>)
Source:	Grown from laboratory culture obtained from Coastal Marine Lab, Hong Kong University of Science and Technology
Size/age:	Log growth phase
Acclimatization:	Grown in 250 mL glass flask (temperature: 22±1°C, salinity: 30‰, 3000 lux) for at least two weeks prior to test.

5.3. Summary of test conditions

Type of test:	Static
Duration:	7 days, 04/08/2014-11/08/2014
Dilution seawater source:	Seawater collected from a pristine site in Clear Water Bay, Sai Kung, Hong Kong
Dilution seawater pretreatment:	Filtered through 0.22 µm membrane
Testing temperature:	22±1 °C
Lighting:	12 h light/12 h dark cycle, 3000±500 lux
Salinity:	30‰
Testing chamber:	Pre-cleaned 100 mL glass beaker
Initial cell density:	(5.0±0.2)×10 ⁴ cell mL ⁻¹
Replicate number:	4
Volume of test medium:	25 mL
Aeration:	None
Reference toxicant:	CdCl ₂
Positive control:	7-day IC ₅₀ toxicity test
Salinity control:	Prepared with ocean salt adding into de-ionized water, salinity: 30‰

5.4. Test results

Table 1. Cell density of diatom *Skeletonema costatum* at the beginning and end of growth inhibition test. Initial cell density: $(5.0 \pm 0.2) \times 10^4$ cell mL⁻¹.

Treatment	Effluent concentration (%)	Cell density after 7-day growth ($\times 10^6$ cell mL ⁻¹)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	1.46	1.25	1.36	1.67	1.44	0.18
Salinity control	0	1.30	1.32	1.25	1.37	1.31	0.05
Concentration 1	2.5	1.63	1.58	1.76	1.73	1.68	0.08
Concentration 2	5	1.89	1.68	1.69	1.80	1.77	0.10
Concentration 3	10	1.85	2.07	2.09	2.13	2.04	0.13
Concentration 4	25	0.75	0.67	0.85	0.86	0.78	0.09
Concentration 5	50	0.14	0.08	0.12	0.14	0.12	0.03

Table 2. Growth rate of *Skeletonema costatum* within 7 days.

Treatment	Effluent concentration (%)	7-day average growth rate (d ⁻¹)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	0.48	0.46	0.47	0.50	0.48	0.02
Salinity control	0	0.47	0.47	0.46	0.47	0.47	0.01
Concentration 1	2.5	0.50	0.49	0.51	0.51	0.50	0.01
Concentration 2	5	0.52	0.50	0.50	0.51	0.51	0.01
Concentration 3	10	0.52	0.53	0.53	0.54	0.53	0.01
Concentration 4	25	0.39	0.37	0.40	0.41	0.39	0.02
Concentration 5	50	0.15	0.18	0.13	0.15	0.15	0.02

5.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during diatom growth inhibition test

Water quality parameters	Effluent concentration (%)						
	Negative control	Salinity control	2.5	5.0	10	25	50
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L ⁻¹)	6.5-13.3	6.6-12.3	6.5-13.1	6.7-12.9	6.4-11.1	6.7-10.3	6.5-10.5
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.7-8.5	7.8-8.3	7.9-8.4	8.0-8.5	8.0-8.4	8.0-8.2	7.7-8.2
Total ammonia (start/end, mg L ⁻¹)	0.08/0.09	0.06/0.06	0.57/0.05	0.89/0.08	1.48/0.22	3.95/1.03	6.45/2.64
Total sulfide (start/end, mg L ⁻¹)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Total residual chlorine (start/end, mg L ⁻¹)	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02
Total suspended solid (mg L ⁻¹)	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2

5.6. IC₅₀ for the diatom *Skeletonema costatum* and test acceptability

Table 4. IC₅₀, none observed effect concentration (NOEC) for the diatom and test acceptability

Parameter	Value	Control limit
Calculated IC ₅₀	(39.7±0.6)%	NA
None observed effect concentration (NOEC)	2.5%	-
Reference toxicant 7-day test:	0.12 mg L ⁻¹	0.13±0.02 mg L ⁻¹
95% of confidence range of reference toxicant test	0.11-0.18 mg L ⁻¹	NA
Temperature variation	<0.5 °C	Average daily temperature variation: ± 1 °C

NA: Not applicable

6. Report on Shrimp Acute Toxicity Test

Test report

6.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove the large debris. Effluent was added with ocean salt in order to raise the salinity to 25‰ and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

6.2. Test organism

Species	Shrimp (<i>Metapenaeus ensis</i>).
Source:	Purchased from local contracted fish dealer
Size/age:	5-7 cm
Acclimatization:	Acclimatized in fully aerated seawater (temperature: 22±1°C, salinity: 25‰) at least 48 hours in the laboratory prior to test. Fed with commercial shrimp feeds.

6.3. Summary of test conditions

Type of test:	Static
Duration:	48 h, 29/07/2014-31/07/2014
Dilution seawater source:	Seawater collected from a pristine site in Clear Water Bay, Sai Kung, Hong Kong
Dilution seawater pretreatment:	Filtered through 0.22 µm membrane
Testing temperature:	22±1 °C
Lighting:	Continuous
Salinity:	25‰
Testing chamber:	Pre-cleaned 20 L tank
Feeding:	None
Number of organisms per replicate:	10
Replicate number:	4
Volume of test medium:	10 L
Aeration:	Moderate, with air stone
Reference toxicant:	CdCl ₂
Positive control:	48 h acute toxicity test
Salinity control:	Prepared with ocean salt adding into de-ionized water, salinity: 25‰

6.4. Test results

Table 1. Survival of shrimps after 48 hours.

Treatment	Effluent concentration (%)	Number of living shrimps after 48 hour (individuals)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	10	10	10	10	10.0	0.0
Salinity control	0	10	10	9	10	9.8	0.5
Concentration 1	6.5	10	9	10	7	9.0	1.4
Concentration 2	12.5	8	10	9	10	9.3	1.0
Concentration 3	25	10	7	8	10	8.8	1.5
Concentration 4	50	8	7	8	9	8.0	0.8
Concentration 5	100	2	4	1	0	1.8	1.7

Table 2. Survival percentage of shrimps after 48 hours.

Treatment	Effluent concentration (%)	Percentage of living shrimps after 48 hour (%)					
		Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	100	100	100	100	100.0	0.0
Salinity control	0	100	100	90	100	97.5	5.0
Concentration 1	6.5	100	90	100	70	90.0	14.1
Concentration 2	12.5	80	100	90	100	92.5	9.6
Concentration 3	25	100	70	80	100	87.5	15.0
Concentration 4	50	80	70	80	90	80.0	8.2
Concentration 5	100	20	40	10	0	17.5	17.1

6.5. Summary of water quality parameters monitoring during test.

Table 3. Summary of water quality parameters during shrimp acute toxicity test.

Water quality parameters	Effluent concentration (%)						
	Negative control	Salinity control	6.5	12.5	25	50	100
Salinity (‰)	25.0	25.0	25.0	25.0	25.0	25.0	25.0
Dissolved oxygen (mg L ⁻¹)	7.5-7.8	7.1-7.7	7.4-7.8	7.4-7.8	7.3-7.8	7.5-7.9	7.4-7.8
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.7-8.0	7.8-8.0	7.7-7.9	7.7-8.0	7.8-8.1	7.7-7.9	7.6-7.8
Total ammonia (start/end, mg L ⁻¹)	0.08-0.15	0.06/0.12	1.03/0.67	1.92/0.85	3.41/2.61	7.62/3.13	12.7/5.73
Total sulfide (start/end, mg L ⁻¹)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
Total residual chlorine (start/end, mg L ⁻¹)	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02	<0.02
Total suspended solid (start/end, mg L ⁻¹)	4.7/5.2	3.8/3.4	5.6/2.9	8.2/4.2	13.5/8.3	20.6/11.6	36.4/15.7

6.6. LC₅₀ for the shrimp *Metapenaeus ensis* and test acceptability

Table 4. LC₅₀ for the *Metapenaeus ensis* and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	(74.2±3.8)%	NA
Negative control survival	97.5%	>90%
Reference toxicant 48-h acute test	5.5 mg L ⁻¹	5.2±0.5 mg L ⁻¹
95% of confidence range of reference toxicant test	4.3-5.3 mg L ⁻¹	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: ±1 °C
Dissolved oxygen concentration	>7.1 mg L ⁻¹	>4 mg L ⁻¹

NA: Not applicable

7. Conclusion

Table 1. Comparison of measured toxicity values with the target toxicity levels.

Test species	Measured LC ₅₀ /IC ₅₀ /NOEC	Target toxicity level
Copepod <i>Tigriopus japonicus</i>	94.7%	≥7.1%
Fish <i>Lutjanus malabaricus</i>	70.6%	≥7.1%
Barnacle larvae <i>Balanus amphitrite</i>	47.2%	≥7.1%
Diatom <i>Skeletonema costatum</i>	39.7% (IC ₅₀)	-
Diatom <i>Skeletonema costatum</i>	2.5% (NOEC)	≥0.51%
Shrimp <i>Metapenaeus ensis</i>	74.2%	≥7.1%

Conclusion: all the measured values met the target toxicity levels as indicated in the EM&A Manual.

Appendix A

Monitoring Data
for
Copepod Acute Toxicity Test

Table 1. Dissolved oxygen concentration and pH in each concentration treatment in copepod acute toxicity test.

Concentration treatment (%)	Dissolved oxygen (mg L ⁻¹)					pH				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	6.8	6.9	7.0	6.8	7.0	7.8	7.9	7.8	8.0	8.0
Salinity control	6.9	7.0	6.7	6.7	7.2	7.8	7.8	7.8	7.8	8.1
6.5	6.7	6.8	6.7	6.9	7.0	7.8	7.9	8.0	8.0	7.9
12.5	6.9	6.8	7.2	7.0	6.7	8.1	7.9	8.0	7.8	8.1
25	7.0	6.7	6.8	6.7	6.8	7.8	7.8	8.0	8.0	8.1
50	6.7	6.7	7.2	7.1	7.0	8.1	7.9	8.0	8.1	8.1
100	7.0	7.2	7.1	7.2	7.0	8.1	8.0	7.9	8.1	7.9

Table 2. Salinity and temperature in each concentration treatment in copepod acute toxicity test.

Concentration treatment (%)	Salinity (‰)					Temperature (°C)				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0

Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the copepod acute toxicity test.

Concentration treatment (%)	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspended solids (mg L ⁻¹)		Total residual chlorine (mg L ⁻¹)	
	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.03	0.13	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
Salinity control	0.07	0.12	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
6.5	0.92	0.83	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
12.5	1.81	2.04	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
25	3.91	4.11	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
50	7.72	7.48	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
100	14.3	13.2	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02

Appendix B

**Monitoring Data
for
Fish Acute Toxicity Test**

Table 1. Dissolved oxygen concentration and pH in each concentration treatment in fish acute toxicity test.

Concentration treatment (%)	Dissolved oxygen (mg L ⁻¹)					pH				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	7.4	7.5	7.7	7.4	7.3	7.9	7.9	8.0	8.0	8.1
Salinity control	7.5	7.7	7.4	7.2	7.5	7.7	8.1	7.9	8.1	7.9
6.5	7.5	7.4	7.5	7.5	7.2	8.2	8.0	7.9	7.9	8.2
12.5	7.7	7.5	7.3	7.3	7.6	8.2	7.9	8.2	8.0	8.1
25	7.2	7.5	7.1	7.5	7.5	8.2	8.2	8.0	8.1	8.2
50	7.5	7.3	7.1	7.4	7.6	8.2	8.1	8.1	8.2	8.2
100	7.3	7.3	7.4	7.3	7.5	7.6	7.5	7.9	7.8	7.7

Table 2. Salinity and temperature in each concentration treatment in fish acute toxicity test.

Concentration treatment (%)	Salinity (‰)					Temperature (°C)				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0

Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the fish acute toxicity test.

Concentration treatment (%)	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspended solids (mg L ⁻¹)		Total residual chlorine (mg L ⁻¹)	
	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.07	0.09	<0.1	<0.1	3.6	4.9	<0.02	<0.02
Salinity control	0.06	0.11	<0.1	<0.1	4.5	5.1	<0.02	<0.02
6.5	1.48	0.87	<0.1	<0.1	6.5	7.5	<0.02	<0.02
12.5	2.31	1.02	<0.1	<0.1	10.1	4.5	<0.02	<0.02
25	4.79	2.35	<0.1	<0.1	15.8	5.7	<0.02	<0.02
50	7.11	3.91	<0.1	<0.1	20.3	8.6	<0.02	<0.02
100	15.1	8.62	<0.1	<0.1	31.2	19.2	<0.02	<0.02

Appendix C

Monitoring Data
for
Barnacle Larvae Acute Toxicity Test

Table 1. Dissolved oxygen concentration and pH in each concentration treatment in barnacle larvae acute toxicity test.

Concentration treatment (%)	Dissolved oxygen (mg L ⁻¹)					pH				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	7.0	6.7	6.7	6.8	6.8	8.0	7.9	8.1	8.0	7.9
Salinity control	6.7	6.6	6.7	6.6	6.5	7.9	7.8	8.0	8.1	7.8
6.5	6.9	6.7	6.9	6.8	6.9	8.1	8.0	8.1	8.0	7.9
12.5	6.8	6.9	6.8	6.7	6.9	7.9	8.1	7.9	8.2	7.8
25	6.7	6.8	6.6	6.8	6.7	8.1	7.9	8.0	7.9	8.1
50	6.7	6.8	6.9	6.7	6.8	7.9	7.8	8.1	7.8	7.9
100	6.8	6.9	6.8	6.8	7.0	7.8	7.7	7.9	7.7	7.6

Table 2. Salinity and temperature in each concentration treatment in barnacle larvae acute toxicity test.

Concentration treatment (%)	Salinity (‰)					Temperature (°C)				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0

Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the barnacle larvae acute toxicity test.

Concentration treatment (%)	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspended solids (mg L ⁻¹)		Total residual chlorine (mg L ⁻¹)	
	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.05	0.07	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
Salinity control	0.09	0.05	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
6.5	0.87	0.65	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
12.5	1.66	1.52	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
25	3.72	3.53	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
50	6.83	6.06	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
100	12.0	11.1	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02

Appendix D

Monitoring Data
for
Diatom Growth Inhibition Test
(Chronic toxicity test)

Table 1. Dissolved oxygen concentration and pH in each concentration treatment in diatom growth inhibition test.

Concentration treatment (%)	Dissolved oxygen (mg L ⁻¹)								pH							
	0 h	24 h	48 h	72 h	96 h	120 h	144 h	168 h	0 h	24 h	48 h	72 h	96 h	120 h	144 h	168 h
Negative control	6.5	6.6	7.4	8.4	10.3	11.5	13.3	9.6	7.7	7.8	7.9	8.2	8.1	8.5	8.1	8.4
Salinity control	6.6	6.8	7.6	8.6	9.9	12.3	10.6	9.6	7.8	8.0	8.0	8.0	8.3	8.3	8.2	8.3
2.5	6.5	6.9	8.1	8.8	9.7	13.1	10.1	10.2	7.9	8.1	8.1	8.2	8.4	8.3	8.3	8.2
5.0	6.7	7.1	7.3	7.8	11.2	12.9	11.5	8.9	8.0	8.2	8.0	8.3	8.5	8.2	8.3	8.5
10	6.8	6.4	6.8	8.1	10.3	9.9	10.5	11.1	8.1	8.0	8.3	8.2	8.0	8.4	8.2	8.2
25	6.9	6.7	6.9	7.9	8.6	9.2	10.3	9.8	7.7	7.8	8.2	8.2	8.1	8.2	8.1	8.0
50	6.5	6.8	6.9	7.4	7.8	9.8	10.4	10.5	7.5	7.7	7.8	7.6	8.2	8.2	8.0	8.1

Table 2. Salinity and temperature in each concentration treatment in diatom growth inhibition test.

Concentration treatment (%)	Salinity (‰)								Temperature (°C)							
	0 h	24 h	48 h	72 h	96 h	120 h	144 h	168 h	0 h	24 h	48 h	72 h	96 h	120 h	144 h	168 h
Negative control	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
2.5	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
5.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
10	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0

Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the diatom growth inhibition toxicity test.

Concentration treatment (%)	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspended solids (mg L ⁻¹)		Total residual chlorine (mg L ⁻¹)	
	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.08	0.09	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
Salinity control	0.06	0.06	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
2.5	0.57	0.05	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
5.0	0.89	0.08	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
10	1.48	0.22	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
25	3.95	1.03	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02
50	6.45	2.64	<0.1	<0.1	<0.2	<0.2	<0.02	<0.02

Appendix E

Monitoring Data
for
Shrimp Acute Toxicity Test

Table 1. Dissolved oxygen concentration and pH in each concentration treatment in shrimp acute toxicity test.

Concentration treatment (%)	Dissolved oxygen (mg L ⁻¹)					pH				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	7.5	7.8	7.8	7.7	7.6	8.0	7.9	7.7	7.7	8.0
Salinity control	7.1	7.5	7.7	7.5	7.5	7.9	8.0	7.8	8.0	7.9
6.5	7.6	7.4	7.8	7.6	7.8	7.7	7.7	7.8	7.8	7.9
12.5	7.5	7.4	7.6	7.8	7.5	7.8	7.7	8.0	7.7	8.0
25	7.7	7.8	7.6	7.3	7.8	7.8	8.1	7.8	7.9	7.8
50	7.8	7.8	7.9	7.7	7.5	7.9	7.8	7.9	7.8	7.7
100	7.8	7.7	7.6	7.4	7.6	7.7	7.6	7.8	7.6	7.8

Table 2. Salinity and temperature in each concentration treatment in shrimp acute toxicity test.

Concentration treatment (%)	Salinity (‰)					Temperature (°C)				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
Salinity control	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
6.5	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
12.5	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
25	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
50	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
100	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0

Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the shrimp acute toxicity test.

Concentration treatment (%)	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspended solids (mg L ⁻¹)		Total residual chlorine (mg L ⁻¹)	
	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.08	0.15	<0.1	<0.1	4.7	5.2	<0.02	<0.02
Salinity control	0.06	0.12	<0.1	<0.1	3.8	3.4	<0.02	<0.02
6.5	1.03	0.67	<0.1	<0.1	5.6	2.9	<0.02	<0.02
12.5	1.92	0.85	<0.1	<0.1	8.2	4.2	<0.02	<0.02
25	3.41	2.61	<0.1	<0.1	13.5	8.3	<0.02	<0.02
50	7.62	3.13	<0.1	<0.1	20.6	11.6	<0.02	<0.02
100	12.7	5.73	<0.1	<0.1	36.4	15.7	<0.02	<0.02